See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/330370206

Phylogeny and global diversity of Porodaedalea, a genus of gymnosperm pathogens in the Hymenochaetales

Article in Mycologia · February 2019



Exploration of Polipora in West Sumatra View project



Mycologia



ISSN: 0027-5514 (Print) 1557-2536 (Online) Journal homepage: http://www.tandfonline.com/loi/umyc20

Phylogeny and global diversity of Porodaedalea, a genus of gymnosperm pathogens in the Hymenochaetales

Fang Wu, Shu-Juan Dai, Josef Vlasák, Viacheslav Spirin & Yu-Cheng Dai

To cite this article: Fang Wu, Shu-Juan Dai, Josef Vlasák, Viacheslav Spirin & Yu-Cheng Dai (2019): Phylogeny and global diversity of Porodaedalea, a genus of gymnosperm pathogens in the Hymenochaetales, Mycologia, DOI: 10.1080/00275514.2018.1526618

To link to this article: https://doi.org/10.1080/00275514.2018.1526618



Published online: 14 Jan 2019.



🖉 Submit your article to this journal 🗗



🌔 View Crossmark data 🗹

Phylogeny and global diversity of *Porodaedalea*, a genus of gymnosperm pathogens in the Hymenochaetales

Fang Wu^{a*}, Shu-Juan Dai^{a*}, Josef Vlasák^b, Viacheslav Spirin ⁶, and Yu-Cheng Dai^a

^aBeijing Advanced Innovation Center for Tree Breeding by Molecular Design, Beijing Forestry University, Beijing 100083, China; ^bBiology Center of the Academy of Sciences of the Czech Republic, Branišovská 31, CZ-37005 České Budějovice, Czech Republic; ^cBotany Unit (Mycology), Finnish Museum of Natural History, P.O. Box 7, FI-00014 University of Helsinki, Finland

ABSTRACT

Porodaedalea is a polypore genus of the Hymenochaetales that encompasses pathogens of conifer trees. In this study, we conduct a comprehensive study of the phylogeny and diversity of *Porodaedalea* based on collections and isolates from Europe, North America, North Africa, and Asia. Phylogenetic analysis of a two-gene data set, nuc ribosomal DNA internal transcribed spacers (ITS1-5.8S-ITS2 = ITS) and translation elongation factor 1-alpha (*tef1*), shows that 20 terminal clades that correspond to phylogenetic species well supported within *Porodaedalea*. Based on morphological evidence, five new species, *P. alpicola*, *P. indica*, *P. kesiyae*, *P. microsperma*, and *P. yunnanensis*, are described and illustrated. In addition, four still unnamed lineages are detected in North America and East Asia.

ARTICLE HISTORY

Received 6 March 2018 Accepted 18 September 2018

Taylor & Francis

Check for updates

Taylor & Francis Group

KEYWORDS

Hymenochaetaceae; phylogeny; taxonomy; wood-inhabiting fungi; 5 new taxa

INTRODUCTION

Porodaedalea Murrill is a small poroid genus of the Hymenochaetales (Basidiomycota) first described over 100 y ago (Murrill 1905). Initially, only P. pini (Brot.) Murrill (the generic type) was recognized in the genus, and subsequently P. chrysoloma (Fr.) Fiasson & Niemelä was separated from P. pini by Donk (1974). For a long time, both species were considered members of Phellinus Quél. and referred to as the Phellinus pini complex (Gilbertson and Ryvarden 1987; Ryvarden and Gilbertson 1994). A more recent circumscription of Porodaedalea was made by Fiasson and Niemelä (1984) on morphological and biochemical grounds, supported by DNA data, and it was found to be a monophyletic group (Wagner and Fischer 2002). Porodaedalea is distinguished from Phellinus by duplex context, generative hyphae bearing crystals at dissepiment edges, long hymenial setae, hyaline to pale yellow and slightly to fairly thick-walled basidiospores, and occurrence on gymnosperm trees.

Porodaedalea is characterized by perennial, effusedreflexed to distinctly pileate to imbricate basidiomes, duplex context of actively growing fructifications, and a dimitic hyphal structure with simple-septate generative and skeletal hyphae. Generative hyphae occasionally produce crystals at dissepiment edges. Setae are abundant, originating mostly from the subhymenium, and

CONTACT Yu-Cheng Dai 🔯 yuchengd@yahoo.com

^{*}Fang Wu and Shu-Juan Dai contributed equally to this paper.

© 2019 The Mycological Society of America

they are rather long and ventricose to subulate. Tramal setae occur in some species, although their presence is not constant and thus of minor taxonomic value. Basidia of *Porodaedalea* are short-clavate, usually distinctly longer than basidioles, and basidiospores are hyaline to pale yellow, slightly to fairly thick-walled, broadly ellipsoid or ovoid to subglobose, and inamyloid but moderately cyanophilous (Murrill 1905; Dai 2010; Tomšovský et al. 2010; Dai et al. 2017). Members of the genus are pathogenic on gymnosperm trees, causing a white pocket rot (Fischer 1994; Dai 2010).

Fourteen species of Porodaedalea are known from all forested continents of the northern hemisphere, although they are more common in the boreal-temperate and conifer-dominated forests. Previous studies reported Porodaedalea from Europe, North America, North Africa, and Asia (Gilbertson and Ryvarden 1987; Fischer 1994; Ryvarden and Gilbertson 1994; Dai 2010; Tomšovský and Kout 2013). In Europe, four species of Porodaedalea are currently recognized: P. pini, P. chrysoloma, P. laricis (Jacz. ex Pilát) Niemelä (= Phellinus abietis (P. Karst.) Jahn (Niemelä et al. 2005; Tomšovský et al. 2010; Ryvarden and Melo 2014), and P. cedrina Pilát ex Tomšovský & Kout, recently described from the Mediterranean area and occurring on Cedrus atlantica (Tomšovský and Kout 2013).

In North America, five described species of Porodaedalea-Porodaedalea piceina (Peck) Niemelä (Niemelä 1985), which belongs to the Porodaedalea "holarctic group" (Brazee and Lindner 2013), P. chrysoloma, P. pini (Gilbertson and Ryvarden 1987), P. cancriformans (M.J. Larsen, Lombard & Aho) T. Wagner & M. Fischer (Wagner and Fischer 2002), and Porodaedalea gilbertsonii (M.J. Larsen) V. Papp (2018)-and two unnamed taxa have been recorded. In addition, Fischer (1994) reported seven unique taxa of Porodaedalea in North America based on mating tests but confirmed only five Porodaedalea groups in the region based on DNA methods (Fischer 1996). Using a multilocus phylogenetic analysis, Brazee and Lindner (2013) indicated that P. pini s. str. and P. chrysoloma s. str. do not occur in North America. Their results were in accordance with earlier studies by Fischer (1994, 1996), Larsen and Melo (1996), Larsen and Stenlid (1999), and Larsen (2000). Only P. piceina and P. cancriformans were mentioned in a list of North American polypores (Zhou et al. 2016).

In Asia, Porodaedalea chrysoloma and P. pini were recognized based on morphology by Núñez and Ryvarden (2000). Dai (2010) recorded five species of Porodaedalea in China: P. chrysoloma, P. himalayensis (Y.C. Dai) Y.C. Dai, P. laricis, P. pini, and P. yamanoi (Imazeki) Y.C. Dai. However, DNA-based taxonomic studies revealed a much more complicated picture. In particular, P. chinensis S.J. Dai & F. Wu was described from southwestern China inhabiting living trees of Pinus yunnanensis (Dai et al. 2017).

Porodaedalea is taxonomically difficult because of the similar morphology among species. Evolutionary relationships of European taxa of *Porodaedalea* were demonstrated by Tomšovský et al. (2010) using phylogenetic analysis based on nuc rDNA internal transcribed spacer (ITS1-5.8S-ITS2 = ITS) and translation elongation factor 1-alpha (*tef1*) sequences, and North American taxa were studied by Brazee and Lindner (2013) with a multilocus phylogeny of four gene regions, including ITS and *tef1*. In Asia, only two species from China were analyzed by molecular data (Tomšovský and Kout 2013; Dai et al. 2017).

In this study, newly collected specimens have been examined by morphological and DNA methods. As a result, five previously unknown species have been detected in Asia. In addition, specimens and sequences of all 20 accepted *Porodaedalea* taxa from the Northern Hemisphere were studied. The aim of this work is to substantiate the current knowledge of phylogeny and species diversity of *Porodaedalea*.

MATERIALS AND METHODS

Morphological studies.—Specimens studied are deposited at BJFC, IFP, H, and in the private herbarium of J. Vlasák (JV), which will be forwarded to PRM. Herbarium abbreviations follow Thiers [continuously updated]. Microscopic procedures follow Dai (2010). At least 30 basidiospores and hymenial setae were measured in each specimen. In presenting basidiospore size data, 5% of the measurements were excluded from each end of the range and shown in parentheses. The following abbreviations are used: IKI = Melzer's reagent, IKI- = negative in Melzer's reagent, KOH = 5% potassium hydroxide, CB = cotton blue, CB+ = cyanophilous, L = mean spore length (arithmetic average of all spores), W = mean spore width (arithmetic average of all spores), Q =variation in the L/W ratios, and n = number of spores measured from given number of specimens. Sections were studied at magnifications up to 1000× using a Nikon Eclipse 80i microscope (Tokyo, Japan) with phasecontrast illumination. Drawings were made with a drawing tube. Color terms follow Petersen (1996).

DNA extraction, PCR, and sequencing.-DNA was extracted from dried basidiomes. ITS and tef1 were selected for genetic analyses. ITS was amplified with the primer pair ITS5 and ITS4 (White et al. 1990), and tef1 sequences were amplified with primers 983F and 2218R (Rehner and Buckley 2005). In some cases, the primer pair 1487F and 2218R was used in nested polymerase chain reaction (PCR) amplifications of tef1 following Brazee and Lindner (2013). The following PCR reaction for ITS was used: initial denaturation at 95 C for 3 min, followed by 35 cycles at 94 C for 40 s, 54 C for 45 s, and 72 C for 1 min, and a final extension of 72 C for 10 min. For tef1, the PCR protocol used a touchdown regime initiated with a 2-min denaturation at 94 C. The annealing temperature in the first amplification cycle was 60 which was subsequently С, incrementally reduced by 1 C per cycle over the next nine cycles. An additional 35 amplification cycles were then performed, each consisting of a 45s denaturation at 94 C, a 90-s annealing step at 53 C, and a 2-min extension at 72 C, concluding with a 10min incubation at 72 C. All newly generated sequences were deposited at GenBank (TABLE 1).

Phylogenetic analyses.—Thirty-four sequences were generated by this study, and 55 related sequences were obtained from GenBank. The ITS and *tef1* data

Table 1. Specimens and GenBank accession numbers of sequences used in this study.

				GenBank accession nos.	
Species	Sample no. Locality Substrate		Substrate	ITS	tef1
Porodaedalea alpicola	Cui 12272 (BJFC)	China, Xizang	Abies sp.	MG585275	MG585294
P. alpicola	Cui 12280 (BJFC)	China, Xizang Abies sp.		MG585276	MG585295
P. alpicola	Cui 12283 (BJFC)	China, Xizang	Abies sp.	MG585277	MG585296
P. cancriformans	1-Sp (CFMR)	USA, California	Abies concolor	JX110042	JX109998
P. cancriformans	FP133112-R (CFMR)	USA, Oregon	Abies magnifica	JX110043	JX109999
P. cedrina	JK 0908/04-3(JV)	Morocco, Middle Atlas	Cedrus atlantica	JQ772468	JQ772474
P. cedrina	MCF 03/1167 (MCF)	Turkey, Fethiye-Babadağ Mt.	Cedrus libani	JQ772467	JQ772473
P. chinensis	Cui 10252 (BJFC)	China, Yunnan	Pinus yunnanensis	KX673606	MG585301
P. chinensis	Dai 15487 (BJFC)	China, Gansu	Gymnosperm	MG585284	MG585302
P. chinensis	Dai 16864 (BJFC)	China, Yunnan	Pinus yunnanensis	KX852282	MG585303
P. chrysoloma	Dai 12674 (BJFC)	Finland, Helsinki	Picea abies	KY000004	MG585306
P. chrysoloma	JV 1411/13 (JV)	Czech, Hluboka	Picea abies	MG585285	MG585305
P. chrysoloma	JV 1408/40-J (JV)	Czech, Hrensko	Picea abies	KY000006	MG585304
P. gilbertsonii	H7002004 (H)	USA, California	Pseudotsuga menziesii	FJ775560	FJ775594
P. gilbertsonii	NAMA 2012-153 (F)	USA, California		MG585290	MG585313
P. gilbertsonii	JV 0709/116-J (JV)	USA, Oregon	Pseudotsuga menziesii	MG585289	MG585312
P. himalayensis	Cui 9320 (BJFC)	China, Xizang	Picea likiangensis	JQ772471	JQ772477
P. himalayensis	Cui 9609 (BJFC)	China, Xizang	Picea likiangensis	MG585286	MG585293
P. himalayensis	Cui 9618 (BJFC)	China, Xizang	Picea likiangensis	KX673604	MG585307
P. himalayensis	Cui 9620 (BJFC)	China, Xizang	Picea likiangensis	KX673605	KX852285
P. indica	Korhonen 1136 (H)	India, Himachal Pradesh	Cedrus deodara	KM011966	_
P. kesiyae	Dai 18417 (BJFC)	Vietnam, Lam Dong	Pinus kesiya	MG585278	MG585297
P. kesiyae	Dai 18427 (BJFC)	Vietnam, Lam Dong	Pinus kesiya	MG585279	MG585298
P. laricis	TFC 1981-38 (TFC)	Russia, Primorsk	Picea ajanensis	FJ775559	FJ775585
P. laricis	H7002007 (H)	Russia, Bashkortostan	Larix sp.	FJ775562	FJ775586
P. microsperma	Cui 12047 (BJFC)	China, Heilongjiang	Larix gmelinii	MG585280	MG585299
P. microsperma	Dai 4926 (BJFC)	China, Inner Mongolia	Picea crassifolia	MG585281	MG585291
P. pini	BRNM 737548 (CFMR)	Turkey, Isparta	Pinus sp.	JQ772470	JQ772476
P. pini	No-6170-T (CFMR)	Portugal, Lisbon	Pinus pinaster	JX110037	JX109993
P. yamanoi	Dai 14795 (BJFC)	China, Jilin	Picea jezoensis	KX673607	MG585309
P. yamanoi	TFC 1971-24 (TFC)	Russia, Sakhalin	Picea jezoensis	FJ775551	FJ775592
P. yamanoi	Dai 8202 (BJFC)	China, Jilin	Picea jezoensis	JQ772469	JQ772475
P. yunnanensis	Dai 3072 (BJFC)	China, Yunnan	Pinus armandii	MG585282	MG585292
P. yunnanensis	Dai 16878 (BJFC)	China, Yunnan	Pinus armandii	MG585283	MG585300
P. sp. 1	FP-103366-T (CFMR)	USA, Georgia	Pinus virginiana	JX110038	JX109994
P. sp. 1	FP-71757 (CFMR)	USA, Virginia	Pinus virginiana	JX110039	JX109995
P. sp. 2	AZ-10-T (CFMR)	USA, Arizona	Pinus strobiformis	JX110040	JX109996
P. sp. 2	AZ-14-T (CFMR)	USA, Arizona	Pinus strobiformis	JX110041	JX109997
P. sp. 3	Miettinen 10543 (H)	China, Jilin	Pinus koraiensis	KM011978	KY024313
P. sp. 3	Spirin 3918 (H)	Russia, Khabarovsk	Larix gmelinii	KM011979	KY024314
P. sp. 4	Spirin 5568-1 (H)	Russia, Khabarovsk	Pinus pumila	KM011968	KY024315
P. sp. 4	Spirin 5567 (H)	Russia, Khabarovsk	Pinus pumila	KM011967	KY024316
P. sp. Holarctic Group 1	FP-135945-T (CFMR)	USA, California	Pseudotsuga menziesii	JX110057	JX110013
P. sp. Holarctic Group 2	FP-97385-T (CFMR)	USA, Colorado	Picea engelmannii	JX110062	JX110018
Onnia leporina	BRNM 712782	Czech, Jihlava	Picea abies	FJ775542	FJ775573

Note. New sequences are in bold.

sets were aligned in BioEdit with Clustal X and edited manually as necessary (Hall 1999). The sequences of *Onnia leporina* (Fr.) H. Jahn were chosen as outgroup following Tomšovský and Kout (2013).

Maximum parsimony (MP) implemented in PAUP* 4.0b10 (Swofford 2003) and Bayesian inference (BI) implemented in MrBayes 3.1.2 (Ronquist and Huelsenbeck 2003) were used in phylogenetic analyses of the individual data sets and the combined data set. Phylogenetic trees were visualized using TreeView. For MP analysis, we used 1000 random sequence additions during a heuristic search, and all characters of the sequences were equally weighted and gaps treated as missing data. MaxTrees was set to 5000, branches of zero length were collapsed, and all parsimonious trees were saved. Branch supports was assessed using a bootstrap (BT) analysis by performing 1000 replicates data sets with random sequence addition for each bootstrap replicate (Felsenstein 1985). BT values >50% are shown. Descriptive tree statistics, tree length (TL), consistency index (CI), retention index (RI), rescaled consistency index (RC), and homoplasy index (HI) were calculated for each tree generated.

The best-fit evolutionary model was determined by MrModeltest 2.3 (Nylander 2004; Posada and Crandall 1998) for the BI analysis. BI was performed using a general time-reversible (GTR) model of DNA substitution and gamma distributed among-site rate variation (Ronquist and Huelsenbeck 2003). Four Markov chains were executed during two independent runs from random starting trees with 4 million generations, keeping one tree every 1000 generations. The first 25% of the samples from the posterior distribution were discarded as the burn-in. A majority rule consensus tree of all remaining trees was calculated. MP and BI analyses generated nearly identical topologies for each data set; thus, only the topology from the MP analysis is presented, along with statistical values from the MP and BI algorithms.

RESULTS

The combined data set had an aligned length of 1784 sites, of which 1483 were constant, 166 variable and parsimony-uninformative, and 135 parsimony-informative. MP analysis yielded four equally parsimonious trees (TL = 415, CI = 0.807, HI = 0.193, RI = 0.854, RC = 0.690). The best-fit model for the *tef1* data set for the BI analysis was a GTR+I+G model. The best-fit models for three partitions of the ITS data set were as follows: HKY+G for ITS1, K80+I+G for 5.8S, HKY+I +G for ITS2. Bayesian analysis resulted in a similar topology as the MP analysis, with an average standard deviation of split frequencies reaching >0.01. The MP tree is provided in FIG. 1. BT and Bayesian posterior probability (BPP) values are shown at internodes.

Asian samples form five well-supported lineages (P. alpicola: BT = 100%, BPP = 1; P. indica: BT =54%, BPP = 0.89; P. kesiyae: BT = 100%, BPP = 1; P. microsperma: BT = 98%, BPP = 1; P. yunnanensis: BT = 100%, BPP = 1), and they are separate from all other known species of Porodaedalea by morphology of basidiomes, the size of hymenial setae, the presence or absence of cystidioles, size and shape of the basidiospores, and preference of plant hosts and distribution. On the basis of the phylogenetic evidence and morphological characters, these species are described below. In addition, four other distinct lineages from North America and boreal East Asia with strong support values were detected: Porodaedalea sp. 1, Porodaedalea sp. 2, Porodaedalea sp. 3 and Porodaedalea sp. 4. These remain undescribed due to unavailable or inadequate collection materials gathered to date.

TAXONOMY

Porodaedalea alpicola S.J. Dai, F. Wu & Y.C. Dai, sp. nov. FIGS. 2A, 3

MycoBank MB823674

Typification: CHINA. XIZANG AUTONOMOUS REGION: Linzhi County, Lulang, Sejila Mountain, 29° 34'24"N, 94°29'10"E, 2850 m, on live *Abies*, 23 Sep 2014, *Cui 12272* (holotype BJFC017186). *Etymology: alpicola* (Latin), referring to the species dwelling in high mountainous areas.

Basidiomes perennial, pileate, solitary to imbricate, without odor or taste, hard corky when fresh, heavy and woody hard when dry. Pilei projecting up to 5 cm, 5.3 cm wide and 0.6 cm thick at base. Pileal surface brown to dull brown, concentrically sulcate with narrow zones, becoming encrusted with age; margin obtuse. Pore surface cinnamon-buff to honey yellow and slightly shining; pores circular to angular, 4–6 per mm; dissepiments thick, entire. Context cinnamonbrown, woody hard, up to 3 mm thick. Tubes cinnamon-buff, woody corky, up to 0.3 cm long.

Hyphal system dimitic, generative hyphae simpleseptate, tissue darkening in KOH.

Context: Generative hyphae hyaline, thin-walled, occasionally branched, frequently simple-septate, 2–3 μ m in diam; skeletal hyphae dominant, yellowish brown, thick-walled with a wide lumen, rarely branched, frequently septate, straight, more or less regularly arranged, 3.5–5 μ m in diam.

Tubes: Generative hyphae frequent, hyaline, thinwalled, occasionally branched, frequently septate, 1.5– 2 µm in diam; skeletal hyphae yellowish brown, thickwalled with a narrow to wide lumen, unbranched, frequently septate, straight, more or less parallel along the tubes, 2.5–3 µm in diam. Hymenial setae frequent, mostly originating from subhymenium, sometimes embedded in trama but not seen at dissepiment edges, subulate, dark brown, thick-walled, encrusted with irregular crystals, $(30-)32-46(-47) \times 5-9(-11)$ µm; cystidioles infrequent, fusoid, hyaline, thin-walled; basidia clavate, with four sterigmata and a simple septum at the base, 11–19 × 5–7 µm; basidioles dominating in hymenium, barrel-shaped to pear-shaped, distinctly shorter than basidia.

Basidiospores $4.5-5.5(-)6 \times 4-4.5(-5) \mu m$, L = 5 μm , W = 4.2 μm , Q = 1.19–1.21 (n = 60/2), broadly ellipsoid to subglobse, hyaline, thin- to slightly thick-walled, smooth, without guttule, usually in tetrads, IKI–, moderately CB+.

Ecology and distribution: on live *Abies*, south Asia, Sep.

Other specimens examined: CHINA. XIZANG AUTONOMOUS REGION: Linzhi County, Lulang, Sejila Mountain, 29°34′24″N, 94°29′10″E, 2850 m, on live Abies smithii, 23 Sep 2014, Cui 12280 (BJFC017194); same locality and date as previous, Cui 12283 (BJFC017197).

Notes: Porodaedalea alpicola is most closely related to *P. himalayensis*; the two species are sister taxa (FIG. 1), which is corroborated by their morphological similarity. However, *P. himalayensis* has an irregularly cracked





Figure 1. MP phylogeny of *Porodaedalea* inferred from the combined ITS and *tef1* sequences. The newly sequenced specimens are in boldface. The tree shown is a phylogram and represents 1 of 10 equally MP trees. Bootstrap and BPP values are indicated at internodes.



Figure 2. Basidiomes of new Porodaedalea species. A. Porodaedalea alpicola (holotype, Cui 12272). B. Porodaedalea kesiyae (paratype, Dai 18417). C. Porodaedalea microsperma (paratype, Dai 4926). D. Porodaedalea yunnanensis (holotype, Dai 16878). Bars = 1 cm.

pileal surface, the generative hyphae at dissepiment edges usually bear crystals, and this species occurs on *Picea* (Dai 2010). In contrast, *P. alpicola* has an uncracked pileal surface, smooth generative hyphae at dissepiment edges, and is so far found on *Abies*.

Porodaedalea indica Spirin, Y.C. Dai & Vlasák, sp. nov. FIG. 4

MycoBank MB823675

Typification: INDIA. HIMACHAL PRADESH: Kullu District, Manali, 32°16'12"N, 77°10'12"E, 2000 m, on live *Cedrus deodara*, 20 Jun 1972, *Korhonen 1136* (holotype H).

Etymology: indica (Latin), referring to India, the country of origin.

Basidiomes pileate, dimidiate, conchate, often with decurrent base, projecting part up to 7 cm wide. Pileal surface dark gray to blackish, indistinctly sulcate, often covered by mosses; margin sharp. Pore surface even to slightly notched, ochraceous brown to rusty-colored when dry; pores angular to sinuous, elongated on sloping substrates, (2–)3–4 per mm, shining; dissepiments rather thin, even to wavy. Context rusty brown, woody

hard, indistinctly fibrillose, about 1 mm thick; tubes indistinctly stratified, ochraceous brown (lower layers) to brown (upper parts), up to 60 mm thick.

Hyphal system dimitic, generative hyphae simpleseptate, tissue darkening in KOH.

Context: Generative hyphae hyaline, thin-walled, occasionally branched, frequently simple-septate, $2-3.5 \mu m$ in diam; skeletal hyphae thick-walled (lumen narrow to capillary), dark brown, regularly arranged, $3.2-4.4 \mu m$ in diam.

Tubes: Generative hyphae hyaline, thin-walled to slightly thick-walled, occasionally branched, frequently simple-septate, 2–3 μ m in diam; skeletal hyphae brown, with a clear lumen, loosely interwoven, 2.3–3.1 μ m in diam. Tramal setae absent. Hymenial setae thick-walled, usually asymmetric, ventricose, some biradicate, mostly with blunt apices, (33–)35–51 × 10–18(–19) μ m; cystidioles common, fusoid, often with long hyphoid apical outgrowth, hyaline, thin-walled; basidia clavate, with four sterigmata and a simple septum at the base, 11–14 × 4–5.5 μ m; basidioles dominating in hymenium, barrel-shaped to pear-shaped, distinctly shorter than basidia.



Figure 3. Microscopic structures of *Porodaedalea alpicola* (holotype, Cui 12272). A. Basidiospores. B. Basidia and basidioles. C. Cystidioles. D. Hymenial setae. E. Hyphae from trama. F. Hyphae from context.

Basidiospores $4-5 \times 4-4.5 \mu m$, L = 4.7, W = 4.2, Q = 1.12 (n = 30/1), subglobose, hyaline, slightly thick-walled, smooth, without guttule, usually in tetrads, IKI–, moderately CB+.

Ecology and distribution: on live *Cedrus*, south Asia (India), Jun.

Notes: Porodaedalea indica is morphologically and phylogenetically close to *P. cedrina* recently described from the Mediterranean (Tomšovský and Kout 2013). Both species occur on living cedar trees, and they share some morphological characters (number of pores per mm, wide, bluntpointed hymenial setae, rather loose tramal tissue). However, hymenial setae are longer ($30-62 \mu m$) and basidiospores larger ($5-6 \times 4-5 \mu m$) in *P. cedrina* in comparison with those of *P. indica* (TABLE 2). In addition, tramal skeletal hyphae of *P. cedrina* ($3.5-5 \mu m$ diam) are wider than in *P. indica* ($2.5-3 \mu m$ diam). *Porodaedalea cedrina* was described based on a single specimen on live *Cedrus* in Himachal Pradesh of India, so its pathogenicity and distribution range is not quite known.

Porodaedalea kesiyae S.J. Dai, F. Wu & Y.C. Dai, sp. nov. FIGS. 2B, 5

MycoBank MB823677



Figure 4. Microscopic structures of *Porodaedalea indica* (holotype, Korhonen 1136). A. Basidiospores. B. Basidia and basidioles. C. Cystidioles. D. Hymenial setae. E. Hyphae from trama. F. Hyphae from context.

Typification: VIETNAM. LAM DONG PROVINCE: Lac Duong District, Bidoup Nui Ba National Park, 12° 07'35"N, 108°33'41"E, 1590 m, on live *Pinus kesiya*, 15 Oct 2017, *Dai 18427* (holotype BJFC).

Etymology: kesiyae (Latin), referring to occurrence on the host tree *Pinus kesiya*.

Basidiomes perennial, pileate, solitary to imbricate, without odor or taste, hard corky when fresh, heavy and woody hard when dry. Pilei projecting up to 4 cm, 6.5 cm wide and 0.5 cm thick at base. Pileal surface grayish brown to black, concentrically sulcate with narrow zones, irregularly cracked and becoming encrusted with age; margin obtuse. Pore surface honey yellow to cinnamon and slightly shining; pores mostly circular to angular, 3–4 per mm; dissepiments thick, entire. Context

dull brown, woody hard, up to 2 mm thick. Tubes cinnamon-brown, woody corky, up to 0.3 cm long.

Hyphal system dimitic, generative hyphae simpleseptate, tissue darkening in KOH.

Context: Generative hyphae hyaline, thin-walled, occasionally branched, frequently simple-septate, 2-3 µm in diam; skeletal hyphae dominant, yellowish brown, thick-walled with a wide lumen, rarely branched, frequently septate, straight, loosely interwoven to more or less regularly arranged, 3-4.5 µm in diam.

Tubes: Generative hyphae frequent, hyaline, thinwalled, occasionally branched, frequently septate, 2-3µm in diam; skeletal hyphae yellowish brown, thickwalled with a narrow to wide lumen, unbranched, frequently septate, straight, loosely interwoven to more or

	Distribution;	Pores				
Species	Substrate	(per mm)	Setae (µm)	Basidiospores (µm)	Cystidioles	Source
P. alpicola	South Asia;Abies	4–6	32–46 × 5–9, L = 36, W = 7	$4.5-5.5 \times 4-4.5,$ L = 5, W = 4.2,	+	This study
P. cancriformans	North America;	3–5	50 × 13	Q = 1.19 - 1.21 4.5-5 × 3.5-4	-	Larsen and Cobb-Poulle (1990)
P. cedrina	North Africa/west Asia; Cedrus	1–2	30–62 × 6–18	5–6 × 4–5,	+	Tomšovský and Kout (2013)
				L = 5.5, W = 4.6, Q = 1.04–1.38		
P. chinensis	Asia; Pinus	2–3	35–69 × 5–14,	4.5–5.5 × 3.5–4.5,	+	This study
			L = 53, W = 9	L = 4.9, W = 4, Q = 1.17–1.26		
P. chrysoloma	Europe; <i>Picea</i>	2–3	30–55 × 6–11,	4–5.5 × 3–4.5,	-	This study
			L = 42, W = 8	L = 5, W = 4.1, Q = 1.24		
P. gilbertsonii P. himalayensis	North America; <i>Pseudotsuga</i> South Asia;	2–3 5–6	30–40 × 7–9 24–43 × 6–10,	5–5.5 × 5 4.5–6 × 3.5–5,	- +	Larsen (2000) This study
	Рісеа		L = 33, W = 8	L = 4.8, W = 4,		
P. indica	South Asia; Cedrus	3–4	35–50 × 10–18,	Q = 1.21 4-5 × 4-4.5	+	This study
			L = 42, W = 14	L = 4.7, W = 4.2, Q = 1.12		
P. kesiyae	Southeast Asia; <i>Pinus</i>	3–4	43–58 × 8–11, L = 50, W = 10	5–5.5 × 4.5–5, L = 5.3, W = 4.6,	_	This study
P. laricis	North Asia;	2–3	30–58 × 6–12,	Q = 1.13 - 1.16 5-5.5 × 3.5-4,	+	This study
	Lunx		L = 45, W = 8	L = 5, W = 3.9, O = 1.27		
P. microsperma	Asia; Larix/Picea	1–2	27–46 × 5–8,	$4-5 \times 3-4$,	-	This study
			L = 35, W = 6	L = 4.3, W = 3.6, Q = 1.17–1.21		
P. pini	Europe; <i>Pinus</i>	1–3	39–67 × 8–15,	5.5–6 × 4–5.5,	+	This study
			L = 51, W = 11	L = 5.9, W = 4.9, Q = 1.2		
P. yamanoi	Northeast Asia; <i>Picea</i>	3–4	36–65 × 7–13,	4–6 × 4–4.5,	+	This study
			L = 48, W = 10	L = 4.9, W = 4.1, Q = 1.19		
P. yunnanensis	South Asia; Pinus	2–3	36-56 × 6-10,	4.5-5.5 × 4-4.5,	+	This study
Demoderately a 2		2.2	L = 49, W = 8	L = 5, W = 4.4, Q = 1.13 - 1.15		This should
Porodaedalea sp. 3 Porodaedalea sp. 4	East Asia; <i>Pinus/Larix</i> East Asia; <i>Pinus</i>	2-3 6-7	33-78 × 9-17 24-38 × 8-12	4.5-5.5 × 3.5-4.5 4.5-5 × 4-4.5	_	This study

 Table 2. Morphological and ecological/geographic features of Porodaedalea taxa.

less parallel along the tubes, 3–4 µm in diam. Hymenial setae frequent, mostly originating from subhymenium, rarely embedded in trama, not seen at dissepiment edges, subulate, dark brown, thick-walled, (30–)43–58(–70) × 8–11(–12) µm; cystidioles absent; basidia clavate, with four sterigmata and a simple septum at the base, 15–22 × 4.5–5 µm; basidioles dominating in hymenium, mostly pear-shaped, distinctly shorter than basidia.

Basidiospores $5-5.5(-6) \times (4-)4.5-5 \mu m$, L = 5.3 μm , W = 4.6 μm , Q = 1.13-1.16 (n = 60/2), subglobose, hyaline, thin- to slightly thick-walled, smooth, without guttule, usually glued in tetrads, IKI-, moderately CB+.

Ecology and distribution: On live *Pinus*, southeast Asia, Oct.

Other specimen examined: VIETNAM. LAM DONG PROVINCE: Lac Duong District, Bidoup Nui Ba National Park, 12°07'35"N, 108°33'41"E, 1590 m, on living tree of *Pinus kesiya*, 15 Oct 2017, *Dai 18417* (BJFC).

Notes: Porodaedalea kesiyae is most closely related to *P. laricis* and two unnamed species from boreal east Asia (FIG. 1). However, *P. laricis* and *Porodaedalea* sp. 3 have slightly bigger pores (2–3 per mm) and a boreal distribution, whereas *P. kesiyae* has smaller pores (3–4 per mm) and occurs in tropical pine forests. *Porodaedalea* sp. 4



Figure 5. Microscopic structures of *Porodaedalea kesiyae* (holotype, Dai 18427). A. Basidiospores. B. Basidia and basidioles. C. Hymenial setae. D. Hyphae from trama. E. Hyphae from context.

differs from *P. kesiyae* in having smaller pores (6–7 per mm), shorter hymenial setae (24–38 μ m), and smaller basidiospores (4.5–5 × 4–4.5 μ m; TABLE 2). *Porodaedalea yunnanensis* and *P. cedrina* have basidiospores similar to those of *P. kesiyae*, but *P. yunnanensis* differs from *P. kesiyae* in having cystidioles, and *P. cedrina* is distinguished from *P. kesiyae* mainly due to the wider hymenial setae (6–18 μ m) and larger pores (1–2 per mm).

Porodaedalea microsperma S.J. Dai, F. Wu & Y.C. Dai, sp. nov. FIGS. 2C, 6

MycoBank MB823678

Typification: CHINA. HEILONGJIANG PROVINCE: Yichun, Xing'an Forest Park, 47°42′23″N, 128°56′41″E, 350 m, on live *Larix gmelinii*, 31 Apr 2014, *Cui 12047* (holotype BJFC016969).

Etymology: microsperma (Greek), referring to the small basidiospores.

Basidiomes perennial, pileate, solitary or imbricate, without odor or taste, hard corky when fresh, heavy and woody hard when dry. Pilei projecting up to 3.8 cm, 5 cm wide and 0.5 cm thick at base. Pileal surface grayish brown to black, concentrically sulcate with narrow zones, irregularly cracked and becoming



Figure 6. Microscopic structures of *Porodaedalea microsperma* (holotype, Cui 12047). A. Basidiospores. B. Basidia and basidioles. C. Hymenial setae. D. Generative hyphae at dissepiment edge. E. Hyphae from trama. F. Hyphae from context.

encrusted with age; margin obtuse. Pore surface claybuff to dull brown and slightly glancing; pores mostly daedaleoid, some circular, 1–2 per mm; dissepiments thick, entire. Context dull brown, woody hard, up to 1 mm thick. Tubes cinnamon, woody corky, up to 0.4 cm long.

Hyphal system dimitic, generative hyphae simpleseptate, tissue darkening in KOH.

Context: Generative hyphae hyaline, thin-walled, occasionally branched, frequently simple-septate, 2–3 μ m in diam; skeletal hyphae dominant, yellowish brown, thick-walled with a wide lumen, rarely branched, frequently septate, straight, more or less regularly arranged, 3–5 μ m in diam.

Tubes: Generative hyphae frequent, hyaline, thinwalled, occasionally branched, frequently septate, some bearing crystals at dissepiment edges, 2–2.5 μ m in diam; skeletal hyphae yellowish brown, thick-walled with a narrow to wide lumen, unbranched, occasionally septate, straight, more or less parallel along the tubes, 2.5–3 μ m in diam. Hymenial setae frequent, mostly originating from subhymenium, not or rarely embedded in trama, not seen at dissepiment edges, subulate, dark brown, thickwalled, encrusted with irregular crystals, (24–)27–46(– 50) × 5–8 μ m; cystidioles absent; basidia clavate, with four sterigmata and a simple septum at the base, 15–17 × 4–5 μ m; basidioles dominating in hymenium, clavate or barrel-shaped, distinctly shorter than basidia. Basidiospores $4-5 \times 3-4 \mu m$, L = 4.3 μm , W = 3.6 μm , Q = 1.17–1.21 (n = 60/2), ovoid, hyaline, thin- to slightly thick-walled, smooth, without guttule, usually in tetrads, IKI–, moderately CB+.

Ecology and distribution: On live *Larix* and fallen *Picea*, Asia, Apr.

Other specimen examined: CHINA. INNER MONGOLIA AUTONOMOUS REGION: Alashan zuoqi (County), Helanshan Natural Reserve, 38°41'22" N, 105°53'19"E, 2150 m on fallen trunk of *Picea*, 29 Apr 2003, *Dai* 4926 (IFP 004120).

Notes: Porodaedalea microsperma is most phenotypically similar to *P. cedrina* described from northern India. However, the latter has longer hymenial setae (30–62 μ m) and larger basidiospores (5–6 × 4–5 μ m) compared with

P. microsperma. Porodaedalea microsperma, P. chrysoloma, P. chinensis, and *P. laricis* have pores of approximately the same size, but *P. chrysoloma* has longer hymenial setae (30–55 μ m) and larger basidiospores (4–5.5 × 3–4.5; TABLE 2). *Porodaedalea chinensis*, shown as the sister lineage to *P. microsperma*, has longer and wider hymenial setae (35–69 × 5–14 μ m) and larger basidiospores (4.5–5.5 × 3.5–4.5 μ m; TABLE 2) compared with *P. microsperma. Porodaedalea laricis* differs from *P. microsperma* by the wider hymenial setae (6–12 μ m) and larger basidiospores (5–5.5 × 3.5–4 μ m; TABLE 2).

Porodaedalea yunnanensis S.J. Dai, F. Wu & Y.C. Dai, sp. nov. FIGS. 2D, 7

MycoBank MB823679



Figure 7. Microscopic structures of *Porodaedalea yunnanensis* (holotype, Dai 16878). A. Basidiospores. B. Basidia and basidioles. C. Cystidioles. D. Hymenial setae. E. Hyphae from trama. F. Hyphae from context.

Typification: CHINA. YUNNAN PROVINCE: Kunming, Heilong Pool, 25°08'34"N, 102°44'52"E, 2460 m, on live *Pinus armandii*, 2 Apr 2016, *Dai 16878* (holotype BJFC022984).

Etymology: yunnanensis (Latin), referring to the locality of the species in Yunnan Province.

Basidiomes perennial, pileate, solitary or imbricate, without odor or taste, hard corky when fresh, heavy and woody hard when dry. Pilei projecting up to 3.3 cm, 5 cm wide and 0.6 cm thick at base. Pileal surface grayish brown to black, concentrically sulcate with narrow zones, irregularly cracked and becoming encrusted with age; margin obtuse. Pore surface buff to cinnamon-buff and slightly glancing; pores circular to angular, 2–3 per mm; dissepiments thick, entire. Context dull brown, woody hard, up to 2 mm thick. Tubes cinnamon, woody corky, up to 0.4 cm long.

Hyphal system dimitic, generative hyphae simpleseptate, tissue darkening in KOH.

Context: Generative hyphae hyaline, thin-walled, occasionally branched, frequently simple-septate, 2–3 μ m in diam; skeletal hyphae dominant, yellowish brown, thick-walled with a wide lumen, rarely branched, frequently septate, straight, more or less regularly arranged, 3.5–5 μ m in diam.

Tubes: Generative hyphae frequent, hyaline, thinwalled, occasionally branched, frequently septate, 1.8– 2.5 µm in diam; skeletal hyphae yellowish brown, thickwalled with a narrow to wide lumen, unbranched, occasionally septate, straight, parallel along the tubes, 2–3.5 µm in diam. Hymenial setae frequent, mostly originating from subhymenium, rarely embedded in trama, not seen at dissepiment edges, subulate, dark brown, thick-walled, $(33-)36-56(-60) \times 6-10(-14)$ µm; cystidioles present, fusoid; basidia clavate, with four sterigmata and a simple septum at the base, $13-20 \times 5-6$ µm; basidioles dominating in hymenium, clavate or barrel-shaped, mostly shorter than basidia.

Basidiospores $4.5-5.5 \times 4-4.5(-5) \mu m$, L = 5 μm , W = 4.4 μm , Q = 1.13-1.15 (n = 60/2), subglobose, hyaline, thin- to slightly thick-walled, smooth, without guttule, usually in tetrads, IKI-, moderately CB+.

Ecology and distribution: On live *Pinus*, south Asia, Apr and Jun.

Other specimen examined: CHINA. YUNNAN PROVINCE: Lijiang County, Yufengshi, 26°59'39"N, 100°11'46"E, 2960 m, on living tree of *Pinus armandii*, 17 Jun 1999, *Dai 3072* (BJFC012865).

Notes: Phylogenetically, *P. yunnanensis* groups with *P. himalayensis* and *P. alpicola* but with poor support (FIG. 1). However, *P. himalayensis* differs from *P. yunnanensis* by smaller pores (5–6 per mm) and shorter hymenial setae (24–43 μ m; TABLE 2). In

turn, *P. alpicola* differs from *P. yunnanensis* by smaller pores (4–6 per mm) and occurrence on *Abies*. *Porodaedalea yunnanensis* is morphologically similar to *P. pini*, and both species inhabit *Pinus*, but *P. pini* has wider hymenial setae (8–15 μ m), usually larger basidiospores (5.3–6.2 × 4–5.5 μ m; TABLE 2), and distribution in Europe.

KEY TO DESCRIBED SPECIES OF PORODAEDALEA

1. Cystidioles present
1'. Cystidioles absent 10
2. Pores 4–6 per mm
2'. Pores 1–4 per mm
3. On Abies P. alpicola
3'. On Picea P. himalayensis
4. On <i>Cedrus</i>
4'. On other gymnosperm trees rather than <i>Cedrus</i> 6
5. Pores 1–2 per mm, basidiospores 5–6 µm long
P. cedrina
5'. Pores 3-4 per mm, basidiospores 4-5 µm long
P. indica
6. On Larix P. laricis
6'. On Picea or Pinus
7. On Picea P. yamanoi
7'. On <i>Pinus</i>
8. Basidiospores 5.5-6 µm long P. pini
8'. Basidiospores 4.5-5.5 μm long
9. Basidiospores broadly ellipsoid, 3.5-4.5 µm wide
P. chinensis
9'. Basidiospores subglobose, 4–4.5 µm wide
P. yunnanensis
10. Pores 1-2 per mm P. microsperma
10'. Pores 2-5 per mm 11
11. Pores 3-5 per mm 12
11'. Pores 2-3 per mm 13
12. On Abies, basidiospores 4.5-5 μm long
P. cancriformans
12'. On <i>Pinus</i> , basidiospores 5–5.5 μm long
P. kesiyae
13. On <i>Picea</i> , basidiospores $<4.5 \mu m$ wide
P. chrysoloma
13'. On <i>Pseudotsuga</i> , basidiospores >4.5 µm wide
P. gilbertsonii

ACKNOWLEDGMENTS

We are grateful to Professor Bao-Kai Cui (BJFC, China) for allowing us to study his specimens and to Dr. Michal Tomšovský (Brno, Czech Republic) for laboratory assistance. The authors thank two anonymous reviewers and an executive editor for comments and suggestions of improvement to an earlier version of the manuscript.

FUNDING

This research was financed by the National Natural Science Foundation of China (project no. 31530002). The research of J. Vlasák was funded by institutional support (RVO 60077344).

ORCID

Viacheslav Spirin D http://orcid.org/0000-0001-5436-6997

LITERATURE CITED

- Brazee NJ, Lindner DL. 2013. Unravelling the *Phellinus pini* s. l. complex in North America: a multilocus phylogeny and differentiation analysis of *Porodaedalea*. Forest Pathology 42:132–143.
- Dai SJ, Vlasák J, Tomšovský M, Wu F. 2017. Porodaedalea chinensis (Hymenochaetaceae, Basidiomycota)—a new polypore from China. Mycosphere 8:986–993.
- Dai YC. 2010. Hymenochaetaceae (Basidiomycota) in China. Fungal Diversity 45:131–343.
- Donk MA. 1974. Checklist of European polypores. Verhandelingen der Koninklijke Akademie van Wetenschappen Afdeeling Natuurkunde Ser. 2(62):1–469.
- Felsenstein J. 1985. Confidence intervals on phylogenetics: an approach using bootstrap. Evolution 39:783–791.
- Fiasson JL, Niemelä T. 1984. The Hymenochaetales: a revision of the European poroid taxa. Karstenia 24:14–28.
- Fischer M. 1994. Pairing tests in the *Phellinus pini* group. Mycologia 86:524–539.
- Fischer M. 1996. Molecular and microscopical studies in the *Phellinus pini* group. Mycologia 88:230–238.
- Gilbertson RL, Ryvarden L. 1987 North American polypores. Oslo, Norway: Fungiflora. 448 p.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/ 98/NT. Nucleic Acids Symposium Series 41:95–98.
- Larsen MJ. 2000. *Phellinus gilbertsonii* sp. nov. from western North America causing heart-rot of coastal Douglas-fir. Folia Cryptogam Estonica 37:51–54.
- Larsen MJ, Cobb-Poulle LA. 1990. *Phellinus* (Hymenochaetaceae). A survey of the world taxa. Synopsis Fungorum 3:1–206.
- Larsen MJ, Melo IM. 1996. Neotypification of *Phellinus pini*. Mycologia 88:839–843.
- Larsen MJ, Stenlid J. 1999. Neotypification of *Phellinus chrysoloma*. Folia Cryptogam Estonica 34:9–13.
- Murrill WA. 1905. The Polyporaceae of North America 11. A synopsis of the brown pileate species. Bulletin of the Torrey Botanical Club 32:353–371.
- Niemelä T. 1985. Mycoflora of Poste-de-la-Baleine, northern Quebec: polypores and the Hymenochaetales. Naturaliste Canada 112:445–472.

- Niemelä T, Kinnunen J, Larsson KH, Schigel DS, Larsson E. 2005. Genus revisions and new combination of some North European polypores. Karstenia 45:75–80.
- Núñez M, Ryvarden L. 2000. East Asian polypores 1. Ganodermataceae and Hymenochaetaceae. Synopsis Fungorum 13:1–168.
- Nylander JAA. 2004. MrModeltest v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University, Uppsala, Sweden.
- Papp V. 2018. Nomenclatural novelties. Index Fungorum. 354:1.
- Petersen JH. 1996. Farvekort. The Danish Mycological Society's colour-chart. Greve, Denmark: Foreningen til Svampekundskabens Fremme.
- Posada D, Crandall KA. 1998. Modeltest: testing the model of DNA substitution. Bioinformatics 14:817–818.
- Rehner SA, Buckley E. 2005. A Beauveria phylogeny inferred from nuclear ITS and EF1-α sequences: evidence for cryptic diversification and links to *Cordyceps teleomorphs*. Mycologia 97:84–98.
- Ronquist F, Huelsenbeck JP. 2003 MRBAYES 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19:1572–1574.
- Ryvarden L, Gilbertson RL. 1994. European polypores 2. Synopsis Fungorum, 6-7:1–743.
- Ryvarden L, Melo I. 2014. Poroid fungi of Europe. Synopsis Fungorum 31:1-455.
- Swofford DL. 2003. PAUP*: Phylogenetic Analysis Using Parsimony. Version 4.0 beta 10. Sunderland, Massachusetts: Sinauer Associates.
- Thiers B. [continuously updated]. Index Herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. [cited 2017 Dec 12]. Available from: http://sweetgum.nybg.org/ science/ih/
- Tomšovský M, Kout J. 2013. *Porodaedalea cedrina* (Basidiomycota, Agaricomycetes, Hymenochaetales)—a new polypore from the Mediterranean area. Nova Hedwigia 96:419–426.
- Tomšovský M, Sedlák P, Jankovský L. 2010. Species recognition and phylogenetic relationships of European *Porodaedalea* (Basidiomycota, Hymenochaetales). Mycological Progress 9:225–233.
- Wagner T, Fischer M. 2002. Proceeding towards a natural classification of the worldwide taxa *Phellinus* s.l. and *Inonotus* s.l., and phylogenetic relationships of allied genera. Mycologia 94:998–1016.
- White TJ, Bruns T, Lee S, Taylor J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ, eds. PCR protocols: a guide to methods and applications. San Diego, California: Academic Press, p. 315–322.
- Zhou LW, Nakasone KK, Burdsall HH Jr, Ginns J, Vlasák J, Miettinen O, Spirin V, Niemelä T, Yuan HS, He SH, Cui BK, Xing JH, Dai YC. 2016. Polypore diversity in North America with an annotated checklist. Mycological Progress 15:771–790.